

Unwinding and rewinding the nucleosome inner turn: Force dependence of the kinetic rate constants

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(Dated: October 15, 2012)

A simple model for the force-dependent unwinding and rewinding rates of the nucleosome inner turn is constructed and quantitatively compared to the results of recent measurements [A. H. Mack *et al.*, *J. Mol. Biol.* **423**, 687 (2012)]. First, a coarse-grained model for the histone-DNA free energy landscape that incorporates both an elastic free energy barrier and specific histone-DNA bonds is developed. Next, a theoretical expression for the rate of transitions across a piecewise linear free energy landscape with multiple minima and maxima is presented. Then, the model free energy landscape, approximated as a piecewise linear function, and the theoretical expression for the transition rates are combined to construct a model for the force-dependent unwinding and re-winding rates of the nucleosome inner turn. Least-mean-squares fitting of the model rates to the rates observed in recent experiments demonstrates that this model is able to well describe the force-dependent unwinding and rewinding rates of the nucleosome inner turn, observed in the recent experiments, except at the highest forces studied, where an additional *ad hoc* term is required to describe the data, which may be interpreted as an indication of an alternate high-force nucleosome disassembly pathway, that bypasses simple unwinding. The good agreement between the measurements and the model at lower forces demonstrates that both specific histone-DNA contacts and an elastic free energy barrier play essential roles for nucleosome winding and unwinding, and quantifies their relative contributions.

I. INTRODUCTION

To fit into a nucleus, eukaryotic DNA is assembled by histones and other proteins into a hierarchy of chromatin structures, and ultimately chromosomes. The fundamental organizational unit of chromatin is the nucleosome in which 146 or 147 bp of DNA are wound around a protein complex comprised of two copies each of the core histones, H2A, H2B, H3 and H4. The core histones are highly conserved across Eukarya, and all have a similar general structure consisting of a central globular domain, an unstructured N-terminal tail, and an unstructured C-terminal tail. The structure of the canonical nucleosome is known in atomic detail as a result of x-ray crystallographic studies [1–3].

By blocking access to promotor DNA, nucleosomes generally repress gene expression in eukaryotes. For highly transcribed genes, the depletion of nucleosomes from the promoter DNA is a key feature and is believed to be a prerequisite for the recruitment of TATA-binding protein and RNA polymerase [4–6]. Nucleosome eviction from the promotor can be accomplished by a number of ATP-dependent chromatin “remodeling” enzymes [7], such as SWI/SNF, which exert force to displace the nucleosomes. It follows that the behavior of nucleosomes under tension, and their mechanical properties more generally, are highly relevant for eukaryotic gene expression. In addition, chromatin is subjected to significant forces during cell division.

Motivated by these considerations, several groups have sought to characterize and understand the forced unwinding

of nucleosomes using optical or magnetic tweezers [8–21]. The accepted model for the nucleosome winding/unwinding pathway, first proposed by Brower-Toland *et al.* [10], is illustrated in Fig. 1. The model envisions four distinct nucleosome states, designated state 2, state 1, state 0, and unbound. State 2 is the canonical nucleosome, in which the histone octamer is wrapped by nearly two turns of DNA. For state 1, the inner turn remains wrapped, but the outer turn is unwrapped. For state 0, both the outer and inner turns are unwrapped, but the histone octamer remains bound to the DNA. Finally, for the unbound state, the histone octamer is dissociated from the DNA.

The purpose of this paper is to present a physics-based model for the force-dependent unwinding and rewinding rates of the nucleosome inner turn, that well describes recent experimentally measured unwinding and rewinding rates, shown in Fig. 2 [22]. To this end, we first develop a simple coarse-grained model for the histone-DNA free energy landscape that incorporates both the elastic free energy barrier described in Refs. 12, 16, and 23 and specific histone-DNA bonds, as indicated in Ref. 19. Secondly, we present a theoretical expression for the rate of transitions across a piecewise linear free energy landscape with multiple minima and maxima. Thirdly, we combine our model free energy landscape, approximated as a piecewise linear function, and our theoretical expression for the transition rates across a piecewise-linear landscape to construct a model for the force-dependent unwinding and re-winding rates of the nucleosome inner turn. Finally, we demonstrate by least-mean-squares fitting that this model provides a good description of the observed force-dependence of the unwinding and rewinding rates of the nucleosome inner turn, as shown as the solid and dashed lines in Fig. 2. At high

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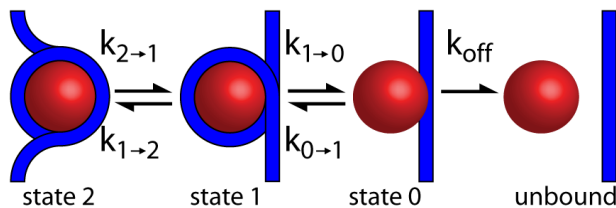


FIG. 1. The four microstates for nucleosome unwinding/rewinding proposed in Ref. 10. Histones are red. DNA is blue. For state 2, the nucleosome is fully wrapped by nearly two turns of DNA. For state 1, the outer turn is unwrapped, but the inner turn is wrapped. For state 0, both the outer and inner turns are unwrapped, but the histone octamer remains bound to the DNA. Finally, the histones may be unbound from the DNA.

forces, an additional *ad hoc* term is required to describe the data, which we interpret as an indication of an alternate high-force nucleosome disassembly pathway, that bypasses simple unwinding. Nevertheless, the good agreement between our measurements and our model at lower forces indicates that both specific histone-DNA contacts and an elastic free energy barrier play essential roles for nucleosome winding and unwinding, and it quantifies their relative contributions.

II. EXPERIMENTAL REVIEW

Using optical tweezers, Brower-Toland *et al.* [10] studied the forced unwinding of nucleosome arrays, assembled from natural avian histones on a sequence of DNA containing 17 tandem repeats of the 5S rRNA nucleosome positioning sequence. They found that under tension a length of DNA equivalent to about 75 bp per nucleosome unwinds apparently continuously at lower forces, corresponding to unwinding of the nucleosome outer turn of DNA. At higher forces, the force-versus-extension curve shows stepwise jumps, also of about 75 bp, each of which corresponds to unwinding the inner turn of DNA. Brower-Toland *et al.* ascribed the observed gradual unwinding of the nucleosome outer turn to relatively weak outer-turn histone-DNA interactions, and the high-force unwinding of the nucleosome inner turn – the transition from state 1 to state 0 – to the sudden disruption of strong histone-DNA interactions, located between 35 and 45 base pairs either side of the nucleosome dyad.

Kulic and Schiessel [12] proposed an alternative explanation for the sudden inner turn disruption events, observed by Brower-Toland *et al.* They showed that the elasticity of a nucleosomal DNA superhelix under tension gives rise to a force-dependent free energy barrier between states with different numbers of turns. This interpretation found support in subsequent experiments, which studied the behavior of avian mononucleosomes under tension, assembled on DNA containing a single 601 nucleosome positioning sequence [16, 17, 23]. Consistent with the predictions of Ref. 12, these experiments reveal that for tensions near 3 pN, the mononucleosome “hops” between states 1 and 2, indicating the exis-

tence of a significant free energy barrier between these states too.

Recently, to further elucidate the energetics of histone-DNA binding, as well as to probe the forces experienced by a polymerase molecule proceeding through nucleosomal DNA during transcription, Hall *et al.* [19] implemented a novel approach, using optical tweezers to unzip double-stranded DNA, wound around a nucleosome, into two single-stranded DNA strands. As the unzipping fork proceeds around the nucleosome, Hall *et al.* found that it dwells at discrete positions, spaced one from another by about 5 bp, indicating the existence of free energy barriers to unzipping, spaced by about 5 bp. Because the dwell times observed were much larger than in the absence of the nucleosome, Hall *et al.* were able to infer that such barriers are the result of discrete histone-DNA binding sites, spaced by approximately 5 bp.

In addition to the 5 bp periodicity, Hall *et al.* also identified three broad regions within each of which the dwell time is especially long, indicating the existence of large free energy barriers to unzipping in these regions. The largest barrier occurs near the nucleosome dyad (region 2), but there are two additional regions in which the barriers are especially high, displaced from the dyad by about ± 45 bp (region 1 near -45 bp and region 3 near $+45$ bp). Barriers are largest in region 2, then comes region 1, with the smallest barriers in region 3. Hall *et al.*’s collected results convincingly demonstrate the existence of significant localized histone-DNA interactions, which should play an essential role in nucleosome unwinding and re-winding, because unwinding necessarily disrupts these bonds.

More recently still, several of us employed optical tweezers in a force-clamp mode to determine, for the first time, the unwinding and rewinding rates of the nucleosome inner turn at several fixed forces (F) [22]. We studied homogeneous nucleosomes containing recombinant wild-type H2A, H2B, H3, and either recombinant wild-type H4 (henceforth H4) or recombinant mutant H4 with arginine 45 mutated to histidine (henceforth H4-R45H), which is an example of a so-called SWI/SNF-independent (Sin) mutation [24–29]. Our motivation for measuring these rates was the observation that the upstream regulatory region (*URSI*) of the yeast *HO* promoter contains a strong nucleosome positioning sequence, which ordinarily requires SWI/SNF for transcription to occur, but that in the absence of SWI/SNF, Sin mutant histones restore transcription from the *HO* locus to wild-type levels. Thus, quantification of the differences between wild-type nucleosomes and nucleosomes containing Sin mutant histones would represent a direct measurement of the minimal free energy change required for an *in vivo* effect, with broad implications for the mechanisms by which histone modifications [30, 31] and histone variations [32–34] affect gene expression. Nucleosomes assembled with H4-R45H also show significantly enhanced thermally-driven nucleosome sliding compared to nucleosomes containing H4 [28].

Our measurements of the unwinding and rewinding rates of the nucleosome inner turn are reproduced in Fig. 2 with the rates displayed on a logarithmic axis, and the corresponding forces displayed on a linear axis. Data for nucleosomes con-

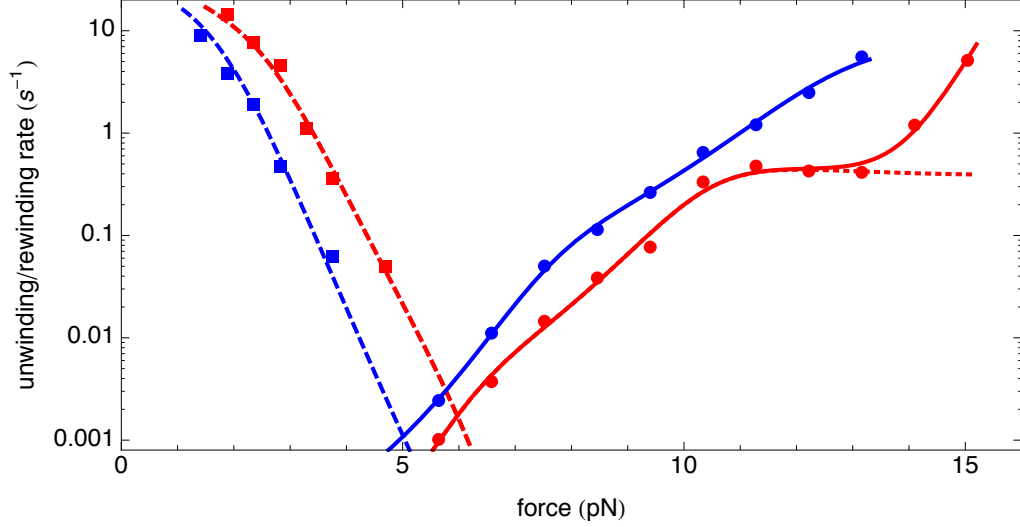


FIG. 2. Comparison of the unwinding and rewinding rates of the nucleosome inner turn for nucleosomes containing H4 (red) or H4-R45H (blue) as a function of force. Circles correspond to the unwinding rate ($k_{1 \rightarrow 0}$). Squares correspond to the re-winding rate ($k_{0 \rightarrow 1}$). The solid and dashed lines are results of least-mean-squares fits to the model described in the text. The red dotted line is the model, but omitting the *ad hoc* term.

taining H4 are shown in red and data for nucleosomes containing H4-R45H are shown in blue. The measured unwinding rates span nearly four orders of magnitude, and the measured rewinding rates more than two orders of magnitude. At each force tested, nucleosomes containing H4-R45H unwind more rapidly and re-wind more slowly than nucleosomes containing H4, indicating that the latter are the more stable. In fact, using the data of Fig. 2, we were able to determine the free energy of the nucleosome inner turn. In brief, there are two contributions to the force-dependent Gibbs free energy difference between state 0 and state 1 (G_{total}), namely the free energy of the nucleosome inner turn ($G_{0 \rightarrow 1}$) and the difference in the Gibbs free energy of the DNA, not wound around the nucleosome [35]. At the force, F^* , at which the unwinding rate and the rewinding rate are equal, state 1 and state 0 have the same Gibbs free energy, and $\Delta G_{total} = 0$. It follows that

$$G_{0 \rightarrow 1} = -F^* d \frac{1 - \sqrt{\frac{k_B T}{F^* L_P}}}{1 - \sqrt{\frac{k_B T}{4F^* L_P}}}, \quad (1)$$

where d is the difference in extension between state 0 and state 1. By extrapolation of the rates in Fig. 2, we find $F^* = 6.0 \pm 0.3$ pN for nucleosomes containing H4, and $F^* = 5.0 \pm 0.3$ pN for nucleosomes containing H4-R45H. It follows, using $L_P \simeq 42$ nm, and $d = 23.3 \pm 0.5$ nm at 5 pN and $d = 23.6 \pm 0.5$ nm at 6 pN that $G_{0 \rightarrow 1} = -142 \pm 7$ pNnm $= -34.6 \pm 1.7 k_B T = -20.8 \pm 1.0$ kcal/mol, and -117 ± 7 pNnm $= -28.5 \pm 1.7 k_B T = -17.2 \pm 1.0$ kcal/mol, for nucleosomes containing H4 and H4-R45H, respectively. The difference in the free energy of the nucleosome inner turn for nucleosomes with H4 and H4-R45H is therefore $6.1 \pm 2.4 k_B T$ [22].

The usual model for how force affects reaction rates, introduced in Ref. 36, predicts an exponential variation of the rates

with force, leading to linear curves, when plotted on semi-logarithmic axes. By contrast, the measured unwinding and rewinding rates, presented in Fig. 2, show significant curvature. Most notable in this regard is that the unwinding rate of nucleosomes containing H4 shows a nearly force-independent rate for forces between about 10 and 14 pN. However, the other three curves also show changes in slope, albeit less dramatic. These deviations from the usual behavior motivate re-consideration of how to appropriately describe these data. (Recent elaborations of Ref. 36 predict curvature in semi-logarithmic plots [37]. However, the predicted curvature is small, only becoming noticeable for rates extending over 7 or 8 orders of magnitude. The changes in slope, we observe, are much larger.)

III. MODEL FREE ENERGY LANDSCAPE OF A NUCLEOSOME UNDER TENSION

A. Elastic free energy of a DNA superhelix under tension

In this section, we present a model for the force-dependent elastic free energy of nucleosomal DNA, treated as an elastic rod, following Ref. [12]. Our calculations are carried out for a single nucleosome, flanked by infinitely-long DNA arms. In comparison, the experiments, that we seek to describe [22], are carried out on 12-nucleosome arrays, held at forces varying from $F = 1.5$ to 15 pN, in which the nucleosomes are in state 1 and are separated from each other by about 40 nm. The characteristic length for elastic deformations of the DNA is given by $\lambda = \sqrt{\kappa/F}$, where κ is the bending modulus of the DNA. Thus, λ varies between 11 nm to 3.4 nm, depending on the force. However, since λ is always many times less than

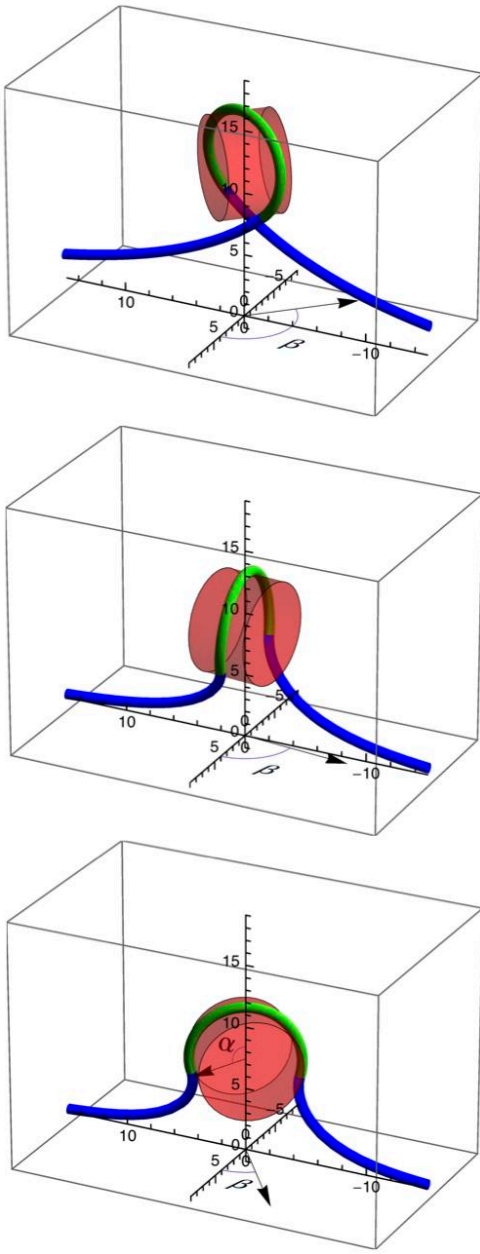


FIG. 3. Elastic-rod models of DNA (blue and green) partially wound around a cylindrical representation of a nucleosome (translucent red) at a force of 8 pN for three different winding (2α) and rotation (β) angles: $2\alpha = 4.8$ radians and $\beta = 2.33$ radians (top); $2\alpha = 4.18$ radians and $\beta = 1.46$ radians (middle); and $2\alpha = 3.7$ radians and $\beta = 0.65$ radians (bottom). DNA in contact with the nucleosome is shown in green. The angle subtended by this DNA defines the winding angle 2α , as indicated in the bottom panel. The rotation angle (β) of the cylinder axis about the vertical axis is indicated in each panel. Axes are marked in nanometers.

the separation between nucleosomes, our treatment, based on isolated nucleosomes, seems reasonable. In addition, we assume torsionally unconstrained DNA, which also corresponds to the experimental situation that we are seeking to describe, because in optical tweezers experiments, the bead, and the

DNA that it is attached to, are free to rotate. Because we treat DNA as an elastic rod, our calculation does not treat the role of different DNA sequences. Our calculations also assume that linker histone H1 is not present, also corresponding to the experimental situation [22].

Illustrated in Fig. 3 are three model configurations of nucleosomal DNA. In each case, the DNA is shown as a blue

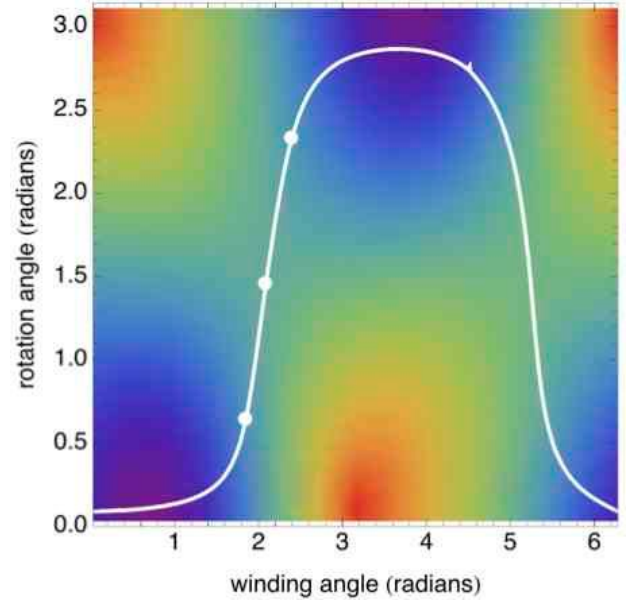


FIG. 4. False color representation of the periodic parts of the elastic free energy at a fixed force of 8 pN, *i.e.* the final two terms in EQ. 2, for which red corresponds to the highest elastic energy and purple the lowest. The white line shows the rotation angle, β , versus winding angle, α , for $F = 8$ pN, according to EQ. 3. The points correspond to the winding angles and rotation angles for the configurations shown in Fig. 3.

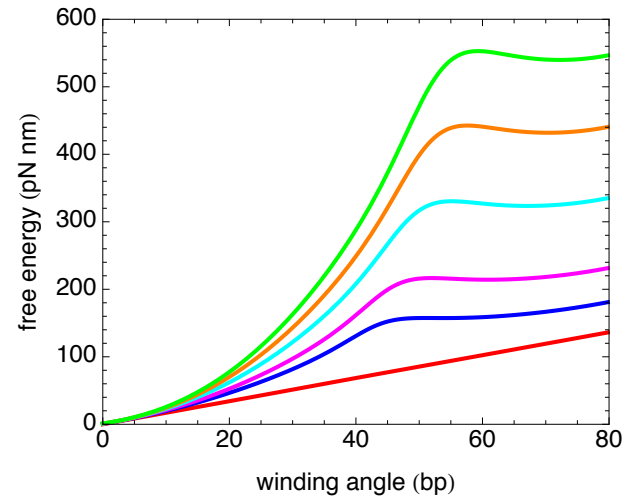


FIG. 5. Elastic free energy landscape for the nucleosome inner turn at zero force (red) at 2 pN (blue), 4 pN (magenta), 8 pN (cyan), 12 pN (orange), and 16 pN (green).

and green rod, partially wound around a histone octamer, represented as a transparent, red cylinder. To model the DNA's elastic energy, we start with the expression, adapted from Ref.

$$\Delta G_{elastic}(F) = \frac{\kappa\alpha}{R} + (2\alpha R - L_C)F \left(1 - \frac{k_B T}{\sqrt{\kappa F}}\right) - 2FR \left(\cos\beta \sin\alpha - \frac{H\alpha \sin\beta}{R}\right) + \sqrt{\kappa F} \left(8 - 4\sqrt{2 + \frac{2R \cos\alpha \cos\beta}{\sqrt{H^2 + R^2}} + \frac{2H \sin\beta}{\sqrt{H^2 + R^2}}}\right). \quad (2)$$

In Eq. 2, κ is the bending modulus of DNA, L_C is its total contour length, R is the superhelical radius, $2\pi H$ is the superhelical pitch, 2α is the winding angle of the DNA about the histone octamer, shown green in Fig. 3, and β is the nucleosome rotation angle. For a nucleosomal superhelix, we take $R = 4.18$ nm, $2\pi H = 2.39$ nm ($H = 0.38$ nm) [1], and $\kappa = 172$ pN nm², corresponding to a DNA persistence length of $L_P = \kappa/(k_B T) = 42$ nm. We routinely find that a value of $L_P = 42$ nm well describes the force-versus-extension of ds-DNA at the solution conditions of our experiments.

The periodic terms in EQ. 2 give rise to a force-dependent

12, for the elastic Gibbs free energy ($\Delta G_{elastic}$) of DNA that is wound into a superhelix, attached to DNA “arms”, and subjected to a tension, F :

In a pulling experiment, the external torque applied to the nucleosome is necessarily zero. This condition implies a relationship among F , κ , α , and β [12], namely

$$F = \frac{2\kappa \left(\sqrt{H^2 + R^2} - P \cos\alpha \cos\beta + H \sin\beta\right) (H \cos\beta + R \cos\alpha \sin\beta)^2}{\sqrt{H^2 + R^2} (H \alpha \cos\beta + R \sin\alpha \sin\beta)^2 \left(R^2 \sin^2\alpha + (H \cos\beta + R \cos\alpha \sin\beta)^2\right)}. \quad (3)$$

It follows that for a given force, F , and winding angle, 2α , the rotation angle, β , is prescribed by Eq. 3. Although an algebraic solution for β is not possible, it is nevertheless straightforward to find the solutions for β from Eq. 3 numerically using Mathematica (Wolfram Research, Urbana, IL). The solution for β , satisfying EQ. 3, corresponding to the lowest free energy path in the $\alpha\beta$ -plane is shown in Fig. 4. Evaluating Eq. 2, using β given by Eq. 3 leads to a one-dimensional free energy landscape, that is a function of the winding angle alone. According to Ref. 1, 126 base pairs (125 base pair spacings, henceforth 125 bp) constitute 1.65 superhelical turns. It follows that there are 75.8 bp per turn. In the remainder of this paper, we have chosen to measure the winding angle of the DNA about the nucleosome in units of base pairs, assuming there are 75.8 bp per superhelical turn. To convert from α in radians to the winding angle in base pairs, it is simply necessary to multiply by $75.8/\pi$.

The one-dimensional elastic free energy landscape for nucleosome winding/unwinding, is plotted versus winding angle for several values of the force in Fig. 5. At each force, the free energy increases super-linearly with increasing winding angle until a local maximum is reached, corresponding to the elas-

tic free energy barrier. Both the value and the location of this maximum increase with force. Beyond the local maximum, the free energy varies only weakly with further increase in the winding angle up to and including values of the winding angle corresponding to state 1 (~ 75 bp). The existence of a free energy barrier, even at large forces, implies that state 1 is mechanically trapped in the low temperature limit.

It is important to emphasize that the elastic energy represented by EQ. 2 is approximate. In particular, the x-ray crystallographically-determined structure of the nucleosome shows significant kinking [1–3]. In addition, recent atomic force microscopy measurements indicate that the elastic energy of highly bent DNA, such as is realized in the nucleosome, is lower than expected on the basis of linear elasticity [38].

B. Specific histone-DNA binding

In addition to the elastic free energy specified in EQ. 2, we must also include histone-DNA binding, which stabilizes the nucleosome. As noted above, the measurements of

Ref. [19] indicate that the histone-DNA interaction shows an approximate 5 bp periodicity, even though the pitch of the DNA double helix is about 10 bp. To explore histone-DNA interactions within the context of the crystallographically-determined nucleosome structure, we have used the program NUCPLOT [39], which automatically identifies protein-DNA interactions, using as input the Protein Data Bank (PDB) file for the protein-DNA complex of interest, *e.g.* the nucleosome. Specifically, we used NUCPLOT, using the default criteria, to identify histone-DNA bonds within the structure of a nucleosome assembled with a modified, palindromic version of 601 DNA, whose structure was recently determined (PDB accession code 3UT9) [3]. A histogram of the number of bonds for each of the DNA strands is shown in Fig. 6. Bonds between histones and DNA in strands I and J are shown in red and blue, respectively. Within each strand, there is an approximate 10 bp periodicity, but the bonds from strand I and from strand J are staggered relative to each other, leading to an overall approximate 5 bp periodicity. Three regions of especially strong bonding, as suggested in Ref. [19], are not apparent from Fig. 6. However, Fig. 6 gives the number of bonds only, and does not factor in bond strength at all, which may be considerably different for different bonds. What Fig. 6 does make clear is that the approximate 5 bp periodicity suggested in Ref. [19] may be understood on the basis of the structure of the nucleosome.

What then is the histone-DNA free energy landscape in the presence of histone-DNA interactions? If there is a strong localized histone-DNA bond at a particular value of the winding angle, then the free energy will decrease in a stepwise fashion when the winding angle is increased beyond that value, cor-

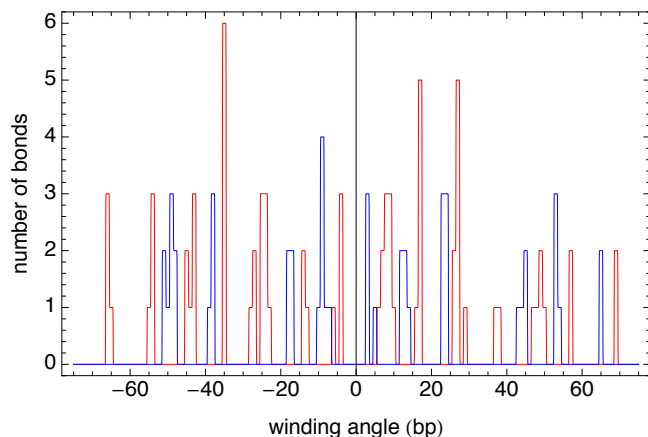


FIG. 6. Histogram of the number of histone-DNA bonds versus winding angle, measured in base pairs from the dyad axis, recognized by NUCPLOT (using its default parameters) for a nucleosome assembled with a palindromic version of 601 DNA (PDB accession code 3UT9) [3]. Shown in red and blue are the bonds between histones and DNA strand I and DNA strand J, respectively. It is important to emphasize that this is the only figure in this paper in which the winding angle is zero at the dyad. In every other figure, zero winding angle corresponds to the point at which no DNA is wound around the nucleosome.

responding to formation of the bond in question. It follows that, if there are strong localized histone-DNA bonds spaced by 5 bp, the histone-DNA free energy landscape as a function of winding angle will show a corresponding sequence of stepwise decreases, as the winding angle is increased, each step separated from its neighbors by 5 bp. Such a simplified free energy landscape, incorporating both the elastic free energy of the DNA superhelix under tension and the effect of histone-DNA binding in a simplified fashion, is illustrated in Fig. 7. As a result of the steps, the net free energy landscape shows multiple minima and maxima as a function of winding angle. Importantly, which local free energy maximum corresponds to the global free energy maximum – *i.e.* to the transition state – changes as a function of force.

C. Transition rates across a piecewise linear free energy landscape

To calculate the unwinding and rewinding rates across a free energy landscape of the sort shown in Fig. 7, we use the “flux over population method” [40], which exploits the observation that the rate in question is equal to the normalized

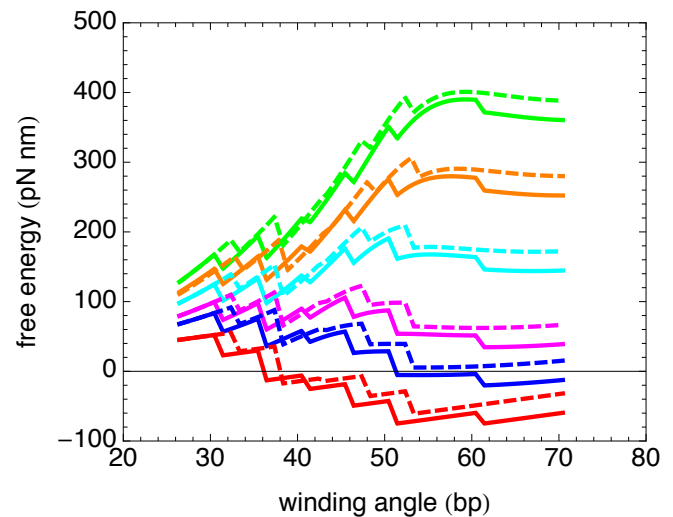


FIG. 7. Model free energy landscapes for the nucleosome inner turn, including the elastic free energy and the free energy of localized histone-DNA binding at 5 bp intervals, at zero force (red) at 2 pN (blue), 4 pN (magenta), 8 pN (cyan), 12 pN (orange), and 16 pN (green). Each step down corresponds to the disruption of one set of histone-DNA bonds. The figure actually illustrates a binding free energy change that occurs over a small range of winding angles (1 bp), corresponding to histone-DNA bonds distributed over this range. The solid curves correspond to the best fit parameters for nucleosomes containing H4, for which the histone-DNA binding near 55 bp and near 65 bp is set to zero, leading to the absence of steps at these locations. The dashed curves correspond to the best fit parameters for nucleosomes containing H4-R45H, for which the histone-DNA binding near 55 bp, 60 bp and near 65 bp is set to zero, leading to the absence of steps at these locations.

steady-state flux into the final state for an adsorbing boundary condition.

First, we consider a discrete model, in which each state along the reaction pathway is labelled by an integer m , with $m = 0$ corresponding to the final state, from which there are no transitions back. Then, the transition rate is equal to the normalized steady-state flux into state 0 for $p_0 = 0$, namely

$$k_{n \rightarrow 0} = p_1 k_{1 \rightarrow 0}. \quad (4)$$

where p_1 is the probability that the system is in state 1, and $k_{1 \rightarrow 0}$ is the rate of transitions from state 1 to state 0. The quan-

tity, $k_{n \rightarrow 0}$, is equal to the inverse of the mean first passage time (MFPT) from state n to state 0 [41]. The probabilities (p_1 , p_2 , *etc.*) are specified in terms of the transition rates between neighboring states, via the steady-state master equations:

$$0 = -(k_{1 \rightarrow 0} + k_{1 \rightarrow 2})p_1 + k_{2 \rightarrow 1}p_2, \quad (5)$$

$$0 = k_{1 \rightarrow 2}p_1 - (k_{2 \rightarrow 1} + k_{2 \rightarrow 3})p_2 + k_{3 \rightarrow 2}p_3, \quad (6)$$

etc. and are normalized: $p_1 + p_2 + \dots + p_n = 1$. It is then straightforward to derive a recursion relation for $k_{n \rightarrow 0}$, namely

$$\frac{1}{k_{n \rightarrow 0}} = \frac{1}{k_{n \rightarrow n-1}} + \left(1 + \frac{k_{n-1 \rightarrow n}}{k_{n \rightarrow n-1}}\right) \frac{1}{k_{n-1 \rightarrow 0}} - \frac{k_{n-1 \rightarrow n}}{k_{n \rightarrow n-1}} \frac{1}{k_{n-2 \rightarrow 0}}, \quad (7)$$

where we take $1/k_{n-2 \rightarrow 0} = 0$ for $n = 2$.

EQ. 7 is useful when the individual transition rates are known. However, if the free energy landscape is a function of the reaction coordinate (*i.e.* the winding angle, as in Fig. 7), it is preferable to employ a continuum description, which permits the transition rates to be determined in terms of parameters that describe the free energy landscape. In the case of a piecewise linear free energy landscape [42, 43], it is possible to derive an analogous recursion formula to EQ. 7, as follows. First, we label the locations of the cusps between neighboring linear regions by an integer m , so that the locations of the cusps are L_m . The difference in free energy between location L_{m+1} (final state) and location L_m (initial state), we designate as $\Delta G_{m \rightarrow m+1}$. Then, similarly to EQ. 4, the transition rate

across a piecewise linear free energy landscape from $x = L_n$ to $x = 0$ is equal to the particle flux at $x = 0$ for adsorbing boundary conditions at $x = 0$, namely

$$k_{n \rightarrow 0} = -D \frac{dp(0)}{dx}, \quad (8)$$

where $p(x)$ is the appropriately normalized probability density, that solves the steady-state Smoluchowski equation, subject to the boundary conditions that $p(0) = 0$ and that $p(x)$ is continuous at the boundaries of each piecewise linear region. (We assume that the free energy is continuous). In this case, we can show that

$$\frac{1}{k_{n \rightarrow 0}} = \sigma_n + \frac{1}{D} \sum_{p=1}^n (L_{p-1} - L_p)^2 \frac{e^{\Delta G_{p \rightarrow p-1}/(k_B T)} - 1 - \Delta G_{p \rightarrow p-1}/(k_B T)}{\Delta G_{p \rightarrow p-1}^2/(k_B T)^2} \quad (9)$$

with

$$\sigma_n = \rho_n + (1 + K_n) \sigma_{n-1} - K_n \sigma_{n-2}, \quad (10)$$

$$K_n = \frac{(-1 + e^{\Delta G_{n \rightarrow n-1}/(k_B T)})(L_n - L_{n-1})/\Delta G_{n \rightarrow n-1}}{(1 - e^{-\Delta G_{n-1 \rightarrow n-2}/(k_B T)})(L_{n-1} - L_{n-2})/\Delta G_{n-1 \rightarrow n-2}}, \quad (11)$$

and

$$\rho_n = - \frac{(L_n - L_{n-1})(L_{n-1} - L_{n-2})(-1 + e^{\Delta G_{n-1 \rightarrow n-2}/(k_B T)})(-1 + e^{\Delta G_{n \rightarrow n-1}/(k_B T)})}{D \Delta G_{n \rightarrow n-1} \Delta G_{n-1 \rightarrow n-2}/(k_B T)^2}. \quad (12)$$

Ordinarily, the second term on the right-hand-side of EQ. 9 is smaller than the first term. Therefore, it may often be a good

approximation to neglect the second term with the result that

$$\frac{1}{k_{n \rightarrow 0}} \simeq \rho_n + (1 + K_n) \frac{1}{k_{n-1 \rightarrow 0}} - K_n \frac{1}{k_{n-2 \rightarrow 0}}, \quad (13)$$

which is formally the same as EQ. 7, provided we identify

$K_n = k_{n-1 \rightarrow n}/k_{n \rightarrow n-1}$, and $\rho_n = 1/k_{n \rightarrow n-1}$. It is also possible

to give an explicit expression for σ_n :

$$\sigma_n = \frac{1}{D} \sum_{p=2}^n \sum_{q=1}^{p-1} \frac{(L_{p-1} - L_p)(L_{q-1} - L_q)(-1 + e^{\Delta G_{p \rightarrow p-1}/(k_B T)}) e^{\sum_{m=q}^{p-2} \Delta G_{m+1 \rightarrow m}/(k_B T)} (-1 + e^{\Delta G_{q \rightarrow q-1}/(k_B T)})}{\Delta G_{p \rightarrow p-1} \Delta G_{q \rightarrow q-1}/(k_B T)^2}. \quad (14)$$

Inspection of EQ. 14 reveals that the MFPT for a landscape with multiple minima and maxima is dominated by the term corresponding to the largest free energy barrier, as expected on the basis of the Arrhenius equation for the transition rate (k) across a free energy barrier, ΔG^\ddagger , namely

$$k = k_0 e^{-\Delta G^\ddagger/(k_B T)}, \quad (15)$$

where k_0 is the rate for zero barrier height ($\Delta G^\ddagger = 0$). However, the factors of $\Delta G_{p \rightarrow p-1}/(k_B T)$ in the denominator of the second term on the right-hand-side of EQ. 9 and in the denominator of the right hand side of EQ. 14 represent significant corrections to the Arrhenius form. Although EQ. 15 is widely-used, it represents an even cruder approximation than EQ. 9.

D. Modelling the nucleosome unwinding and rewinding rates

Although derived for a piecewise linear free energy landscape, EQ. 9 through EQ. 12 express the transition rates solely in terms of free energy differences and reaction coordinate differences. Therefore, these equations may be readily applied to a general free energy landscape. This approach is equivalent to approximating the landscape in question as a piecewise linear landscape, and is the approach we have taken to model the winding and unwinding rates for the nucleosome inner turn. Specifically, to create a minimal model free energy landscape for the nucleosome inner turn at each force, we add together the elastic energy given by EQ. 2 and a set of linear steps, one for each histone-DNA binding location. For the step near base pair m , we take the free energy to decrease linearly from base pair $m + \delta - 1/2$ to base pair $m + \delta + 1/2$, where m is a positive integer and δ specifies the registration between the zero of the winding angle and the zero of the 5 bp periodicity.

Fig. 7 is an example of such a model landscape. In the following, it is convenient to denote the location of base pair $m + \delta - 1/2$ as location m and the location of base pair $m + \delta + 1/2$ as location $m.5$, and to denote the change in binding free energy from initial location m to final location $m.5$ as ΔG_m . To apply EQ. 9 through EQ. 12, we identify successive locations appearing in these equations with locations 25.5, 30, 30.5, 35, 35.5 *etc.* Thus, we calculate the unwinding and rewinding rates across the model free energy landscape.

To fit the model rates so-obtained to the data shown in Fig. 2, we are lead to introduce the following possible fitting parameters: δ , which specifies the registration between the zero of the winding angle and the 5 bp periodicity; D , which is an effective rotational diffusion coefficient; and a set

of histone-DNA binding energies (ΔG_m). To limit the number of fitting parameters to as few as possible, we held fixed the DNA persistence length ($L_p = 42$ nm), the spacing between histone-DNA binding sites (5 bp), and the width of each histone-DNA binding site (1 bp).

In both state 1 and state 0, the number of base pairs of DNA in contact with the histone octamer is not precisely known. However, to numerically calculate the transition rates between state 1 to state 0, it is necessary to pick definite starting and ending winding angles. For the purposes of our calculation of the transition from state 1 (0) to state 0 (1), therefore, we assume a starting winding angle corresponding to base pair number 70 (25) and an ending winding angle corresponding to base pair number 25 (70), which correspond to the free energy landscape shown in Fig. 7. In fact, EQs. 9 through 14 show that the transition rate is a sum of terms, and that the rate is determined by the largest free energy barrier encountered, *i.e.* the largest term in EQ. 14. It follows therefore that even if the starting and ending base pairs are actually larger and smaller, respectively, than 70 and 25, provided there is not a significant contribution to the free energy barrier from base pairs between the actual starting and ending base pairs and base pairs 70 and 25, respectively, the calculated model rates will be essentially unchanged. Our assumption that 70 and 25 are reasonable end points may be justified *a posteriori* on the basis of the success of the fits, we achieve. The histone-DNA binding energies within the included region, which are therefore possible fitting parameters, are: ΔG_{30} , ΔG_{35} , ΔG_{40} , ΔG_{45} , ΔG_{50} , ΔG_{55} , ΔG_{60} , and ΔG_{65} . We assume that each of these binding energies is negative, corresponding to binding.

Irrespective of the values of these fitting parameters, it is not possible to achieve satisfactory agreement between our model and the two experimental data points obtained at high forces ($F > 14$ pN) for nucleosomes containing H4. Therefore, we have added to our model rate an *ad hoc* term, $D \exp[\Delta G^\ddagger + F x^\ddagger/(k_B T)]$ [36], which is negligible at low forces, but which is able to match these two data points. In principle, such a term describes a process that is an alternative to unwinding, and competes with unwinding. For example, we may speculate that a high-force alternative to unwinding is that the DNA may slide sideways off of the nucleosome.

Because the values of the rates span several decades, in order to achieve satisfactory agreement between the measured rates and our model over the full range, we carried out least-mean-squares fits of the logarithm of the model rates to the logarithm of the measured rates, using Mathematica's `NonLinearModelFit` function. We sought to limit, as far as possible, the total number of fitting parameters, while still achieving a satisfactory description of our experimental data

(Fig. 2). Therefore, we fitted all of our rate-constant data simultaneously, so that the unwinding and rewinding rates were both described by the same set of binding free energies. Furthermore, we used the same effective rotational diffusion constant for unwinding and rewinding and for nucleosomes containing H4 and nucleosomes containing H4-R45H. To further restrict the number of fitting parameters, we fixed ΔG_{40} by requiring that the free energy of a nucleosome with H4 at $F = F^* = 6.0$ pN is the same at locations 25.5 and 70, and that the free energy of a nucleosome containing H4-R45H at $F = F^* = 5.0$ pN is the same at locations 25.5 and 70, as found by extrapolating our experimental measurements. Finally, because initial fitting revealed that, for nucleosomes containing H4, the rates could be well-described with $\Delta G_{55} = 0$ and $\Delta G_{65} = 0$, and, for nucleosomes containing H4-R45, the rates could be well-described with $\Delta G_{55} = 0$, $\Delta G_{60} = 0$ and $\Delta G_{65} = 0$, these parameters were subsequently held fixed at zero. The resultant best fits are shown as the solid and dashed lines in Fig. 2, including the *ad hoc* term for the unwinding of nucleosomes containing H4. Evidently, the model rates provide a good description of our measurements. The corresponding best fit parameters are shown Table I.

IV. DISCUSSION

A. Mapping landscape to histone-DNA binding sites

It is important to realize that how to relate the binding locations across the free energy landscape to specific positions within the nucleosome is not straightforward in the absence of additional information or assumptions. This is because each 5 bp of DNA between successive binding locations can detach

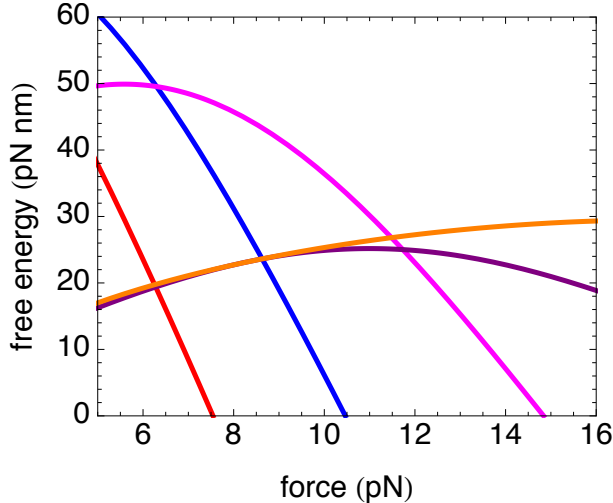


FIG. 8. Free energy differences between location 70 and location 40 (red line), location 45 (blue line), location 50 (magenta line), location 55 (purple line), and the location of the elastic free energy maximum (orange line) versus force.

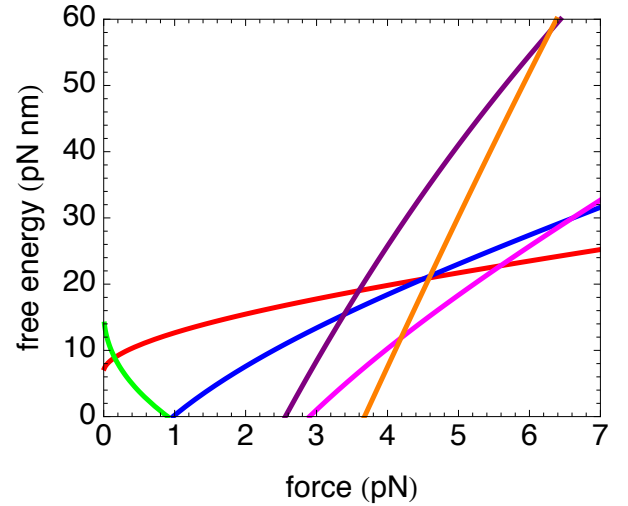


FIG. 9. Free energy differences between location 25 and location 30 (red line), location 35 (blue line), location 40 (magenta line), location 45 (purple line), and location 50 (orange line) versus force. Also plotted is the free energy difference between location 25 and location 70 (green line) versus force, which represents the largest barrier at the smallest forces.

from either side of the histone octamer, leading to the same reduction in the winding angle.

B. Mapping landscape to rates

To understand which features of our model free energy landscape determine the unwinding and rewinding rates and how well each of the fitting parameters is determined, it is instructive to plot the difference in free energy between the starting location (location 14 for unwinding and location 5.5 for rewinding) and various intermediate locations on the unwinding or the rewinding pathway. Examples of such free energy differences are shown using the best fit parameters for nucleosomes containing H4 in Fig. 8 (unwinding) and Fig. 9 (rewinding). Each of these curves represents a free energy barrier to unwinding at the force in question. We may expect the largest free energy barrier to unwinding at a particular force to be rate limiting at that force. Clearly, for forces up to about 6.5 pN, the rate-limiting barrier to unwinding is at location 9; for forces between about 6.5 and 11.5 pN, the rate-limiting barrier to unwinding is at location 10; for forces above about 11.5 pN, the rate-limiting barrier to unwinding is at the location of the elastic free energy maximum.

In the context of our free energy landscape model, the maximum barrier height for unwinding at the elastic free energy maximum depends on ΔG_{60} ; at location 10, it depends on $\Delta G_{50} + \Delta G_{50}$; and, at location 9, it depends on $\Delta G_{45} + \Delta G_{50} + \Delta G_{60}$. Therefore, we may expect each of these parameters to be well-determined by fitting the unwinding rates. Importantly, for forces above 11.5 pN, the barrier to unwinding actually increases only slightly with increas-

	H4	H4-R45H
ΔG_{30}	-30.9 ± 30.6 pN nm ($-7.5 \pm 7.5 k_B T$)	-28.4 ± 9.9 pN nm ($-6.9 \pm 2.4 k_B T$)
ΔG_{35}	-44.4 ± 34.1 pN nm ($-10.8 \pm 8.3 k_B T$)	-54.6 ± 7.7 pN nm ($-13.3 \pm 1.9 k_B T$)
ΔG_{40}^{***}	-20.9 pN nm ($-5.1 \pm k_B T$)	-4.8 pN nm ($-1.2 k_B T$)
ΔG_{45}	-32.5 ± 1.8 pN nm ($-7.9 \pm 0.4 k_B T$)	-30.3 ± 1.2 pN nm ($-7.4 \pm 0.3 k_B T$)
ΔG_{50}	-34.0 ± 2.6 pN nm ($-8.3 \pm 0.6 k_B T$)	-33.8 ± 3.3 pN nm ($-8.2 \pm 0.8 k_B T$)
ΔG_{55}	0.0 pN nm*	0.0 pN nm*
ΔG_{60}	-16.9 ± 1.3 pN nm ($-4.1 \pm 0.3 k_B T$)	0.0 pN nm*
ΔG_{65}	0.0 pN nm*	0.0 pN nm*
δ	0.95 ± 0.41 bp	2.85 ± 0.52 bp
D	5820 ± 1340 bp ² s ^{-1**}	$5820 \pm 1340 \pm$ bp ² s ^{-1**}
ΔG^\ddagger	-135 ± 30 pN nm ($32.9 \pm 7.3 k_B T$)	N.A.
x^\ddagger	8.8 ± 2.0 nm	N.A.

TABLE I. Best fit parameters that yield the model rates (lines) in Fig. 2. *Parameters held fixed at zero. **Rates for nucleosomes containing H4 and nucleosomes containing H4-R45H were fit simultaneously using the same value of D for both data sets. ***Fixed to ensure that the unwinding and rewinding rates are equal at F^* . The quoted errors correspond to the standard errors produced by Mathematica's NonLinearModelFit via the ParameterErrors property.

ing force. This feature of the model naturally accounts for the experimental observation that the unwinding rate is nearly force-independent within the force range between 11.5 and 14 pN. Thus, we assert that this feature of our data is especially convincing evidence that a correct description indeed involves the elastic free energy barrier in an essential fashion. The corresponding plot for nucleosomes containing H4-R45H (not shown) reveals that within the range of forces studied experimentally the rate-limiting barrier is either at location 9 or at location 10. This is a result of $\Delta G_{60} = 0$ for nucleosomes containing H4-R45H.

For rewinding, Fig. 9 reveals that for forces less than about 3.5 pN, location 6 corresponds to the free energy barrier to rewinding, and that for forces between about 3.5 and 6.5 pN location 9 corresponds to the free energy barrier to rewinding. The height of the barrier at location 6 is independent of any of our binding energy parameters. The height of the barrier at location 9 involves the sum $\Delta G_{30} + \Delta G_{35} + \Delta G_{40}$. Therefore, we may expect this sum to be well-determined by our fits to the rewinding rate. However, we should expect the individual component free energies to be less well-determined by the fits. This observation explains why the fitting errors are relatively large for ΔG_{30} and ΔG_{35} in comparison to the fitting errors for ΔG_{45} , ΔG_{50} , ΔG_{55} , and ΔG_{60} .

C. Alternative models

We may inquire how well our rate measurements and model are able to discriminate against alternative models of histone-DNA binding. For example, a number of authors have postulated a constant histone-DNA binding free energy per bp [12, 16, 23, 44], albeit often with different values for the inner and outer turns. Accordingly, we have calculated the force-dependent rates expected in the case of a constant histone-

DNA binding free energy per base pair of 4.1 pNnm per bp, or equivalently 12 pN. We picked a value of 12 pN in order to ensure that the free energy for a winding angle of 25 bp is equal to the free energy for a winding angle of 70 bp at a force of 6 pN. These calculations are shown as the green solid (unwinding) and dashed (rewinding) lines in Fig. 10. The corresponding free energy landscape is shown in Fig. 11. Also reproduced in this figure are the measured and model unwinding and rewinding rates for nucleosomes containing H4, which are shown as the solid and dashed red lines, and the red circles and squares. The constant-binding-energy-per-bp model predicts rates with significantly different force-dependences than observed experimentally and reproduced by our model. In addition, the unwinding rate is a factor of about 100-fold slower than observed. Thus, our data and modeling rules out such a model. We also tested models that impose a 10 bp spacing between histone-DNA binding sites, but were unable to achieve satisfactory fits with such a model (not shown).

D. Effective rotational diffusion constant

According to our fitting results, the effective rotational diffusion coefficient of the nucleosome is $5800 \text{ bp}^2 \text{ s}^{-1}$. In comparison, the rotational diffusion coefficient of a sphere of radius $R = 4.2 \text{ nm}$ is given by

$$D^* = \frac{k_B T}{8\pi\eta R^3} = 2 \times 10^6 \text{ s}^{-1} \simeq 2.9 \times 10^8 \text{ bp}^2 \text{ s}^{-1} \quad (16)$$

where $\eta = 0.001 \text{ Pas}$ is the viscosity of the fluid in which the sphere rotates. Therefore, there is a factor 5×10^4 discrepancy between our measured value of D and the value appropriate for free rotation of a sphere of nucleosomal radius. How then does our best-fit value for the effective diffusion coefficient compare to results in the literature? Via fluorescence experiments, Li *et al.* measured the rewinding rate for 27 bp of the

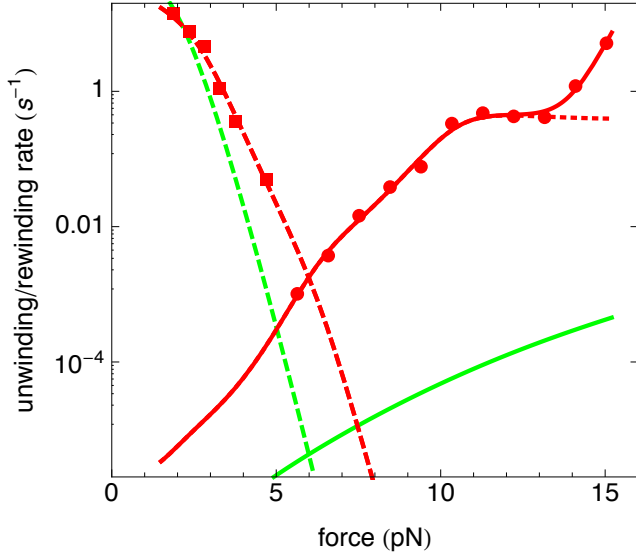


FIG. 10. Comparison of the unwinding and rewinding rates of the nucleosome inner turn, according to the best-fit local interaction model of histone-DNA interactions (dashed and solid red lines), and a model that posits a uniform binding free energy per unit length (dashed and solid green lines). Red circles correspond to the measured unwinding rate ($k_{1 \rightarrow 0}$), and red squares correspond to the measured re-winding rate ($k_{0 \rightarrow 1}$) for nucleosomes containing H4. The red dotted line is the local interaction model, but omitting the *ad hoc* term. The green curves correspond to a constant histone-DNA binding energy per bp, chosen so that at a force of 6 pN the free energies of 25 and 70 bp wound are equal.

nucleosome outer turn to lie between 20 and 90 s^{-1} at zero force [45]. Based on our measured rotational diffusion coefficient for the nucleosome inner turn, and assuming a flat coarse-grained free energy landscape, we would predict a rate to rewrap 27 bp of $5800/27^2 = 8 s^{-1}$ close to the experimental result of Ref. 45. In fact, for a free energy landscape that is inclined downwards, as a result of histone-DNA binding, we may expect somewhat faster rewinding and even better agreement. Our results for D also seem consistent with the extrapolations to zero force of the nucleosome *outer turn* rewinding rates given in Refs. [16] and [17] of about 100 s^{-1} in both cases.

To explain the apparent discrepancy between our result for the rotational diffusion coefficient and EQ. 16, we turn to Refs. 46 and 47, which show theoretically that long-length-scale diffusion across a free energy landscape with short-scale “roughness” may be described on the basis of a coarse-grained free energy landscape with an effective diffusion coefficient that is renormalized by a factor that accounts for the short-scale roughness. Refs. 46 and 47 are directly applicable here: The free energy landscape shown in Fig. 7 is by construction a coarse-grained free energy, since the molecular details of the histone-DNA interaction are omitted. It follows that the diffusion coefficient that emerges from consideration of such a landscape is necessarily a renormalized diffusion coefficient.

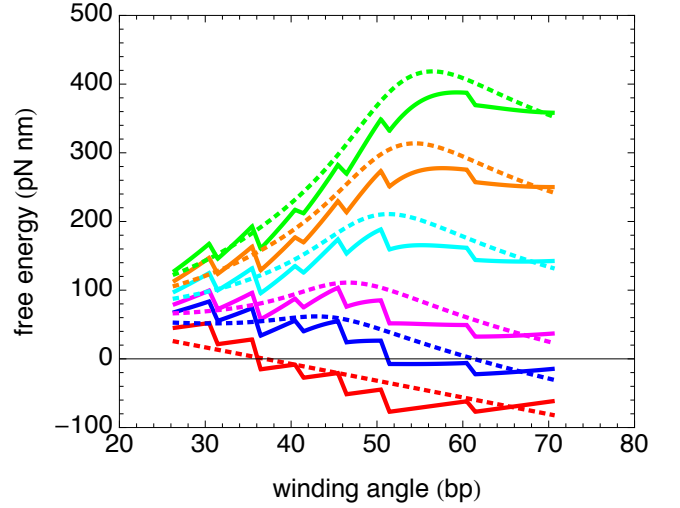


FIG. 11. Model free energy landscapes for the nucleosome inner turn at zero force (red) at 2 pN (blue), 4 pN (magenta), 8 pN (cyan), 12 pN (orange), and 16 pN (green). The solid curves reproduce those shown in Fig. 7, for which each step down corresponds to the disruption of one set of histone-DNA bonds occurring over a small range of winding angles equivalent to 1 bp. The dotted curves correspond to a constant histone-DNA binding energy per bp, chosen so that at a force of 6 pN the free energies of 25 and 70 bp wound are equal.

Because diffusion and friction are related via the Einstein relation, a renormalized diffusion coefficient may also be interpreted in terms of internal friction [48].

Our best fit value of $D = 5800 \pm 1300 \text{ bp}^2 \text{ s}^{-1}$ is the effective rotational diffusion coefficient, corresponding to the coarse-grained histone-DNA interaction free energy of Fig. 7. If we assume that the molecular-scale diffusion coefficient is $D^* = 5 \times 10^7 \text{ rads}^2 \text{ s}^{-1}$, then we have $D/D^* = 5 \times 10^{-6}$. To facilitate comparisons with the results of Refs. 48 and 49, we estimate the “roughness” of the histone-DNA interaction at the scale of individual bonds via the expression for Gaussian landscape roughness, given in Ref. 46, namely $\Delta G = k_B T \sqrt{\log(D^*/D)}$. Thus, we find $\Delta G \simeq 3.5 k_B T$. In comparison, Ref. 49 determines a roughness of $5.7 k_B T$ for the forced unbinding of the GTPase Ran from the nuclear transport receptor importin- β , and Ref. 48 determines a roughness of $4 k_B T$ for stretching cellulose and dextran. Thus, our results are in-line with these studies, as well as with Ref. 45. In fact, the reaction rates in single molecule experiments are generally orders of magnitude smaller than naive expectations. Presumably, this is because free energy landscape roughness is ubiquitous.

E. Asymmetric unwinding model

Why are the unwinding and rewinding rates of nucleosomes containing H4 different from those of nucleosomes containing H4-R45H? To answer this question, we turn to the histone-DNA interaction map presented in Ref. [19]. In the context

of the nucleosome inner turn, which contains a total of about 76 bp, inspection of the interaction map indicates that, if the nucleosome inner turn were to consist of DNA that is wound symmetrically ± 38 bp from the dyad axis of the histone octamer, then significant portions of regions 1 and 3 would both largely lie beyond the nucleosome inner turn. On the other hand, if the nucleosome inner turn is asymmetric with respect to the dyad axis and consists of base pair -55 to base 21, for example, relative to the dyad, then all or almost all of the binding energy from both region 1 and region 2 stabilizes the nucleosome inner turn. In this scenario, region 3 is involved solely in stabilizing the nucleosome outer turn. We therefore hypothesize, as shown schematically in Fig. 12, that the nucleosome under tension in state 1 realizes an asymmetric configuration, because of its lower free energy compared to the symmetric configuration. That an asymmetric partially-unwound nucleosome configuration could be realized at zero force was suggested previously in Ref. [50].

According to this hypothesis, in order to unwind the nucleosome inner turn, it is necessary to break the strong histone-DNA bonds in either region 1 or region 2. If the binding in region 1 is weaker than in region 2 [19], we may expect unwinding to occur via breaking region 1 bonds. We hypothesize that this is the situation in the case of nucleosomes containing H4. According to this scenario, there would be a more-or-less linear relationship between binding location and position within the nucleosome, with the nucleosome dyad located approximately near a winding angle of about 12 bp, so that the strong bonding near the dyad spans winding angles from 0 to about 25 bp, and the landscape described by our fitting parameters for nucleosomes containing H4 corresponds to region 1. This hypothetical situation is illustrated in the bottom panel of Fig. 13. By contrast, for nucleosomes containing H4-R45H, we suppose that the binding energy in region 2 is reduced, so that it is now smaller than the binding energy in region 1. As a result, in this scenario, unwinding occurs by breaking region 2 bonds, and the landscape described by our fitting parameters in the case of nucleosomes with H4-R45H corresponds to region 2. This hypothetical situation is illustrated in the top panel of Fig. 13. According to this hypothesis, the bonds disrupted in the transition from state 1 to state 0 are different for nucleosomes containing H4 than for nucleosomes containing H4-R45H. Therefore, as a result, we may expect that the force-dependent rate for unwinding the nucleosome inner turn of nucleosomes containing H4-R45H will be different than that for nucleosomes containing H4, as we observe experimentally. Furthermore, on this basis, we predict that nucleosomes containing H4 and nucleosomes containing H4-R45H show different dissociation rates to the unbound state. However, since unwinding and rewinding the nucleosome outer turn does not involve region 2 bonds, we also predict that the winding and unwinding rates of these nucleosomes' outer turn are the same for nucleosomes containing H4 and nucleosomes containing H4-R45H.

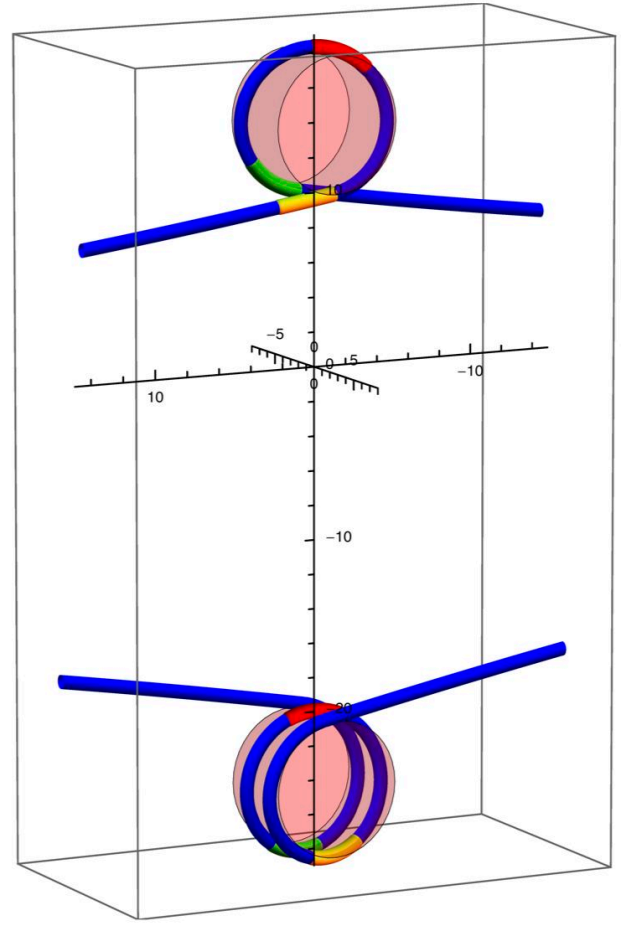


FIG. 12. Schematic of the DNA configuration in nucleosome state 2 (depicted as 147 bp) held at a tension of 2 pN (bottom) and nucleosome state 1 (depicted as 75 bp) held at a tension of 8 pN (top), according to our model. The three regions of DNA that form strong histone-DNA bonds in the canonical nucleosome structure (state 2), according to Ref. [19], are shown green (region 1), red (dyad region/region 2), and yellow (region 3). In the figure, in state 2, the DNA is depicted symmetrically arranged about the center of nucleosome dyad (red). However, in state 1, in which about 75 base pairs of DNA are wound around the histone octamer, because regions 1 and 3 are separated by more than about 75 base pairs (see Fig. 1 of Ref. [19]), histone-DNA bonds in only two of the three regions of strong histone-DNA binding – either regions 1 and 2 or regions 2 and 3 – can be simultaneously satisfied. This is indicated in the figure by the yellow DNA that was bound to the histone octamer in state 2, being not in contact with the histone octamer, and therefore not bound, in state 1. According to this hypothesis, symmetry about the nucleosome dyad is not preserved in state 1, as illustrated in the figure. The axes are marked in nanometers.

V. CONCLUSIONS

Building on the experiments presented in Ref. [22], the principal result of this paper is that the force-dependent unwinding and rewinding rates of the nucleosome inner turn can be explained (except at the highest forces) on the basis of a simple, physical model that incorporates in an essen-

tial fashion both the elastic free energy barrier to unwinding and rewinding introduced in Ref. [12] and elaborated in Refs. [16, 23], and localized histone-DNA binding with an approximate 5 bp periodicity, as proposed in Ref. [19]. This analysis provides new insight into nucleosome winding and unwinding, the energetics of histone-DNA interactions and will be important for the growing numbers of simulations of nucleosome and chromatin behavior that are appearing in the literature [51–63]. Beyond an improved understanding of nucleosome unwinding and rewinding, these results also have important implications for theories of nucleosome sliding [44, 64–67], other processes that involve partial or complete unwinding of dsDNA from the histone octamer [7], and the mechanisms of eukaryotic gene expression more generally [4–6].

ACKNOWLEDGMENTS

We thank J. Antonypillai, C. Cheng, and R. Ilgen, for valuable discussions. This work was supported by the Raymond

and Beverly Sackler Institute for Biological, Physical and Engineering Sciences and NSF PoLS 1019147. D.J.S. is the recipient of a NSF Graduate Research Fellowship.

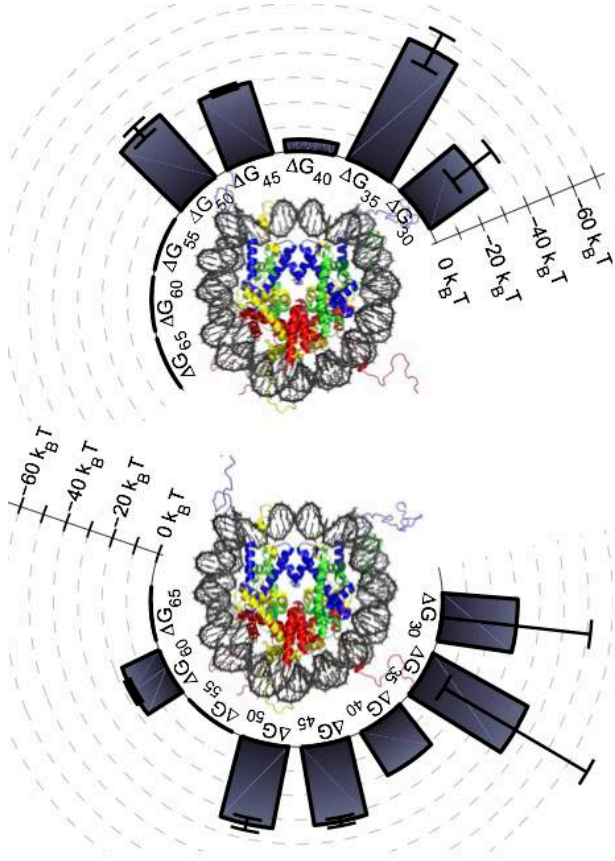


FIG. 13. Hypothetical mapping of binding energies to nucleosome locations. Bottom: Hypothetical mapping of bond energies to nucleosome locations for nucleosomes containing H4. Top: Hypothetical mapping of bond energies to nucleosome locations for nucleosomes containing H4-R45H.

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